

## OK! Kit

The easy way to check the performance of Thermal Cyclers

### Protocol

- Transfer 10  $\mu$ l of OK! Mix (Tube 1) into the PCR tube
- Add 10  $\mu$ l DNA/Primers (Tube 2).
- Overlay with mineral oil if necessary
- Place in the Thermal Cycler

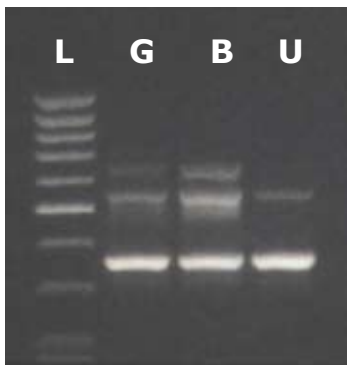
If possible, set the Thermal Cycler to ramp at medium rate (1.2 to 2.5°C/sec)

### Cycling profile

Initial denaturation step: 95°C for 5 mins  
Then cycle 31 times:  
Step 1: 95°C for 30 secs  
Step 2: 66°C for 60 secs  
Step 3: 72°C for 60 secs

After cycling, load 10  $\mu$ l onto 1.7% agarose gel and electrophorese alongside a 100 bp DNA ladder

Expected fragment sizes: 650 bp (top), 550 bp (middle) & 360 bp (bottom)



### Interpretation of results

L = 100 bp DNA ladder

G = Good machine: - The 3 fragments are all present, but in different intensity  
Top band just visible. Machine working correctly

B = Bad machine: - Top band almost as strong as the middle band  
Machine cooling below the required annealing temp.

U = Ugly machine: - Faint middle band and top band fainter still (may even not be visible)  
Machine not reaching to the required annealing temp.

### Related Products

**100 bp DNA ladder** = An agarose size standard supplied  
'ready to load' in a room temp. stable loading buffer

Supplied as: Tube 1 (Blue Amplification Mix) & Tube 2 (Primers/DNA Mix) in multiple of 1 ml aliquots  
Store at -20°C

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